# Role of Regenerative Methods in Management of Amputation Stump

Nishad K<sup>1</sup>, Neljo Thomas<sup>2</sup>, Ravi Kumar Chittoria<sup>3</sup>, Barath Kumar Singh<sup>4</sup>, Jacob Antony Chakiath<sup>5</sup>

How to cite this article: Nishad K, Neljo Thomas, Ravi Kumar Chittoria, *et al.*/Role of Regenerative Methods in Management of Amputation Stump/J Orth. Edu. 2023;9(1):85–88.

#### Abstract

Wound healing is an array of carefully planned steps playing in a sequence to mettle the ill fate that nature has brought on the body. It includes inflammation, proliferation and remodelling. Wound healing depends on different growth factors, cytokines, interleukins and cell population for success. In this article we report how we have used different regenerative methods like the autologous platelet rich plasma, insulin therapy, prolotherapy, low level laser therapy for augmenting in wound healing.

Keywords: Platelet rich plasma; Regenerative medicine; Wound healing.

## INTRODUCTION

Wound healing can become delayed due to multitude of reasons like infection, foreign material, lack of growth factors etc. the wound healing can be augmented by using different methods like growth factors, NPWT. The wound healing can also be delayed by the large size of the wound and surface area of the wound that needs

Author Affiliation: <sup>1,2,4-5</sup>Senior Resident, <sup>3</sup>Professor, Department of Plastic Surgery, Jawaharlal Institute of Postgraduate Medical Education and Research, Pondicherry 605006, India.

**Corresponding Author: Ravi Kumar Chittoria**, Professor, Department of Plastic Surgery, Jawaharlal Institute of Postgraduate Medical Education and Research, Pondicherry 605006, India.

E-mail: drchittoria@yahoo.com

Received on: 13.12.2022

Accepted on: 31.12.2022

to be covered wound that are covered by skin graft also needs to be supported with different growth factors.

#### MATERIALS AND METHODS

The patient is a 42 year male manual labourer who had a road traffic accident and sustained injury to his right lower limb. he underwent above knee amputation of the lower limb and sustained surgical site infection. The wound had to be open and was decided for secondary healing. However, the repeated dressing, antibiotics to mitigate infection could not bring healing to the raw area (Fig. 1). Hence, we resorted to use regenerative methods to supplement the growth factors. We used methods of platelet rich plasma (Fig. 2), prolotherapy (Fig. 3), Low level laser therapy (Fig. 4) for wound bed preparation according to the TIME principle. We also used non cultured keratinocyte suspension for the wound (Fig. 5), dermal pixel grafting (Fig. 6) to supply growth factors to the raw area. The wound bed was prepared and skin grafting (Fig. 7) was done to cover the raw area. The skin graft was also



Fig. 1: Raw area at presentatio





Fig. 2: Aprp fowoud Bed Preparation

supplemented with regenerative methods (Fig. 8).



Fig. 3: Prolotherapy For Wbp



Fig. 4: Lowlevel Laser Therapy



Fig. 5: No Cultured Keratinocyte Graft



Fig. 6: Dermal Pixel Graft



Fig. 8: LLLT to skin graft area

## DISCUSSION

Platelet Rich Plasma (PRP) is a biological product defined as a portion of the plasma fraction of autologous blood with platelet concentration above the baseline (before centrifugation).<sup>1</sup> PRP contains high levels of platelets and also the full complement of clotting factors, the latter remaining at their normal, physiologic levels.<sup>2</sup> It is comprised of a range of growth factors, chemokines, cytokines, and other plasma proteins.<sup>3</sup> PRP is a source of signaling molecules, and upon activation of platelets in PRP, the P-granules degranulate



Fig. 7: Skin Grafting done with Donor Site



Fig. 9: Healed Wound

and release GFs and cytokines that will change the pericellular micro-environment. Some of the most important GFs released by platelets in PRP include vascular endothelial GF(VEGF), fibroblast GF (FGF), platelet derived GF(PDGF), epidermal GF, hepatocyte GF, insulin like GF1,2 (IGF-1, IGF-2), matrix metalloproteinases (MMP)2,9, and interleukin 8.<sup>4,5</sup>

LLLT, phototherapy or photo biomodulation refers to the use of photons at a non-thermal irradiance to alter biological activity.<sup>6</sup> LLLT at low doses has been shown to enhance cell proliferation of fibroblasts<sup>7-10</sup>, keratinocytes<sup>11</sup>, endothelial cells<sup>12</sup> and lymphocytes.<sup>13,14</sup>

In vitro studies have shown that cultivation of cells in high glucose culture medium can increase the PDGF expression. PDGF has multiple reparative effects in wounds, including promotion of angiogenesis, fibroblast proliferation, and extracellular production. TGF-b expression is also increased by high glucose.<sup>15,16</sup> TGF-b is involved in different steps of wound healing from inflammation to wound re-epithelialization. Other growth factors increased by high glucose include EGF, b-FGF, IGF and CTGF.

## CONCLUSION

We have used different regenerative methods to augment the wound healing process and have found it to be useful. However, it needs large randomised control trials to be used in large scale. The limitation of the study was that it was done in a single patient.

# DECLARATIONS

## Acknowledgment

*Authors' contributions:* All authors made contributions to the article.

Availability of data and materials: Not applicable.

Financial support and sponsorship: None.

Conflicts of interest: None.

*Consent for publication:* Not applicable.

# REFERENCES

- 1. Alves R, Grimalt R: A randomized placebocontrolled, double-blind, half-head study to assess the efficacy of platelet-rich plasma on the treatment of androgenetic alopecia. Dermatol Surg 2016;42:491–497.
- Wroblewski AP, Melia HJ, Wright VJ: Application of platelet-rich plasma to enhance tissue repair. Oper Tech Orthop 2010;20:98–105.
- 3. Lynch MD, Bashir S: Applications of plateletrich plasma in dermatology: a critical appraisal of the literature. J Dermatolog Treat 2016; 27:285–289.
- Andia I, Abate M: Platelet rich plasma: underlying biology and clinical correlates. Regen Med 2013; 8:645–658
- 5. Ferrando J, Fernández-Sartorio C, González de

Cossío AC, Navarra E: Tratamiento de la alopecia androgenetica con factores de crecimiento plaquetario. Monografías Dermatología 2016; 42:491–497.

- Avci P, Gupta A, Sadasivam M, Vecchio D, Pam Z, Pam N, Hamblin MR. Low-level laser (light) therapy (LLLT) in skin: stimulating, healing, restoring. InSeminars in cutaneous medicine and surgery 2013 Mar (Vol. 32, No. 1, p. 41). NIH Public Access.
- Lubart R, Wollman Y, Friedmann H, Rochkind S, Laulicht I. Effects of visible and near infrared lasers on cell cultures. J Photochem Photobiol B. 1992;12(3):305–310. [PubMed] [Google Scholar]
- Wu W, Naim JO, Lanzafame RJ. The effect of laser irrafiation on the release of bFGF from 3T3 fibroblasts. Photochem Photobiol. 1994;59(2):167– 170. [PubMed] [Google Scholar]
- 9. Vinck EM, Cagnie BJ, Cornelissen MJ, Declercq HA, Cambier DC. Increased fibroblast proliferation induced by light emitting diode and low power laser irradiation. Lasers Med Sci. 2003;18(2):95–99. [PubMed] [Google Scholar]
- Frigo L, Fávero GM, Lima HJ, Maria DA, Bjordal JM, et al. Low-level laser irradiation (InGaAIP-660 nm) increases fibroblast cell proliferation and reduces cell death in a dose-dependent manner. Photomed Laser Surg. 2010;28(Suppl 1):S151– S156. [PubMed] [Google Scholar]
- Basso FG, Oliveira CF, Kurachi C, Hebling J, Costa CA. Biostimulatory effect of low-level laser therapy on keratinocytes in vitro. Lasers Med Sci. 2013;28(2):367–374. [PubMed] [Google Scholar]
- 12. Szymanska J, Goralczyk K, Klawe JJ, Lukowicz M, Michalska M, et al. Phototherapy with low-level laser influences the proliferation of endothelial cells and vascular endothelial growth factor and transforming growth factor-beta secretion. J Physiol Pharmacol. 2013;64(3):387–391.
- 13. Moore P, Ridgway TD, Higbee RG, Howard EW, Lucroy MD. Effect of wavelength on low-intensity laser irradiation-stimulated cell proliferation in vitro. Lasers Surg Med. 2005;36(1):8–12.
- Agaiby AD, Ghali LR, Wilson R, Dyson M. Laser modulation of angiogenic factor production by T-lymphocytes. Lasers Surg Med. 2000;26(4):357– 363.
- 15. Penn JW, Grobbelaar AO, Rolfe KJ. The role of the TGF-beta family in wound healing, burns and scarring:a review. Int J Burns Trauma 2012;2:18–28
- 16. Freeman JW, Empson YM, Ekwueme EC, Paynter DM, Brolinson PG. Effect of prolotherapy on cellular proliferation and collagen deposition inMC3T3-E1 and patellar tendon fibroblast populations. Transl Res 2011; 158:132–139

000%000<sup>.</sup>