

Occurrence of Antibiotic Residues in Milk: Detection and Public Health Concerns

Abhishek Sharma¹, Atul Kumar², Neelam Sharma³

How to cite this article:

Abhishek Sharma, Atul Kumar, Neelam Sharma/Occurrence of Antibiotic Residues in Milk: Detection and Public Health Concerns/ Journal of Animal Feed Science and Technology 2022;10(1):23-29.

Abstract

There is an increase in demand for food of animal origin, due to increasing global population. Therefore, livestock owners around the globe are trying to increase animal product output which ultimately leads to enormous production pressure on the livestock. To achieve such high production goals, animals should be disease free and physiologically fit. To ensure their healthy status, antibiotics are being used extensively in therapeutic & sub-therapeutic doses to enhance production, which ultimately lead to presence of antibiotic residues in the final animal products like milk, meat, eggs etc. If such contaminated products are consumed by humans for a longer period of time, it may subsequently lead to significant health and economic issues ranging from emergence of AMR to starter failure in dairy industry. Therefore, present review focuses on various aspects of antibiotic usage in animal husbandry practices, initiative taken to combat AMR, methods used for detection of antibiotics in milk and their impact on human health.

Keywords: Antibiotic residues; Animal products; Detection methods; Human health risks.

Introduction

Animal sourced foods such as milk, meat, eggs, fish, and honey are being consumed by people across globe. They are considered nutritious and essential for growth and development of human

body. But their image of being nutritious and healthy products is now getting tarnished. This is due to occurrence of pathogenic microorganism, antibiotics, pesticides, mycotoxins, hormones, heavy metals etc. in animal feed and animal sourced foods especially from India.¹⁻⁹ Among various xenobiotic, antibiotics are more commonly reported in animal sourced foods. Antibiotics are the substances which are produced naturally by living organisms or synthetically in laboratory conditions, and are able to kill or inhibit the growth of other microorganisms. Antibiotics enters the human body directly or indirectly by using antibiotics as growth promoters, feed efficiency enhancers or for treatment of various infectious diseases in animals.¹⁰ These are one of the most commonly used veterinary medicine in relation to animal feed production and around 80% of animals used in food production are treated with antibiotics. The current global average consumption of antibiotics per kg of animal produced is >100mg/kg. Present use of antimicrobials for animal production in India is 30 kg/km² which is expected to grow 312% by

Author Affiliation: ¹Post Graduate Research Scholar, ²Assistant Professor, ^{1,2}Department of Veterinary Public Health and Epidemiology, ³Professor and Head, Department of Chemistry and Biochemistry, Chaudhary Sarwan Kumar Himachal Pradesh Agricultural University, Palampur 176062, Himachal Pradesh, India.

Corresponding Author: Atul Kumar, Assistant Professor, Department of Veterinary Public Health and Epidemiology, Chaudhary Sarwan Kumar Himachal Pradesh Agricultural University, Palampur 176062, Himachal Pradesh, India.

E-mail: dratul9@gmail.com

Received on: 12.11.2021

Accepted on: 12.12.2021

2030.¹¹

In 1940s, the growth promoter effect of antibiotics was discovered when it was observed that animals show improved growth after feeding on dried mycelia of *Streptomyces auriofaciens*. Use of antibiotics as growth promoter may improve feed efficiency by 17% in cattle, 10% in lambs, 15% in poultry & 15% in swine.¹² Certain antibiotics are also used as premixes (Premixes are the mixtures of one or more active ingredients of antibiotic compounds that are exclusively used for preparation of medicated veterinary feeds). Medicated veterinary feeds are fed to animals after being mixed with animal feed. Examples of such antibiotics in medicated feed include oxytetracycline @ 4.4%, amoxicillin & ampicillin @ 0.025% owing to their high antimicrobial property and ability to resist digestion by gastric juices.

Antibiotics are administered to animals through various routes like parenteral, oral or topical. The residues level of antibiotics in animal produce are highest in the animals which are treated via injectable routes in comparison to oral routes. These antibiotic residues eventually appear in animal produce such as milk, meat and eggs. After consumption by the consumers through contaminated animal sourced foods, these residues are responsible for causing adverse health effects in humans such as carcinogenicity (oxytetracycline, furazolidone), nephropathy (gentamycin), mutagenicity, hepatotoxicity, reproductive disorders, anaphylactic shock (penicillins) and even transfer of antibiotic resistant bacteria (ARB) or antibiotic resistant gene (ARG) to humans.¹²⁻¹³ In current world scenario, antimicrobial resistance has been a concerning problem and challenging towards treatment of various diseases. Apart from health effects on consumers, antibiotic residues in milk may interfere with fermentation process resulting in starter failure. This then contributes to significant economic losses in the dairy industry.

In context of above mentioned facts, the present review focuses on antibiotics usage in animal husbandry practices, their impact on public health and methods used for detection of antibiotics residues in milk.

Material and methods

The information presented in this review was obtained from a literature search of free electronic databases such as Google Scholar, PubMed, and Science Direct with regard to publications

on antibiotic residues in milk. The key words used were “antibiotic”, “antibiotic residues”, “antibiotic residues in milk”, “antibiotic residues in feed”, “antimicrobial drug residues in food”, “antimicrobial residues”, “antimicrobial drugs in animals”, “drug residues in milk”, “antimicrobial drug residues in milk”. The exclusion criteria included incomplete publications (i.e. papers with abstract only) and papers published in non-indexed journals. The inclusion criteria included full length original research papers, review articles, short communications in indexed and peer-reviewed journals.

Reasons for occurrence of antibiotics residues in animal sourced foods and food safety regulations in India Since there is ever increasing demand for milk & meat products, therefore antimicrobials have been an irreplaceable part of animal health care sector worldwide. For production of better quality products, the primary focus is always on the quality of raw products. Therefore, the health and disease free status of producing animal is a must from economic as well as welfare point of view. Antibiotics have been one of the most common medicine used for treatment of wide variety of infections in both humans and animals. However, residues of these drugs when excreted from body of treated animals or secreted in animal produce, may directly or indirectly enters the food chain. Direct sources of antibiotics to animals include parenteral or oral administration of antibiotic drugs, fodder mixed with antibiotics to enhance feed efficiency, intramammary or intrauterine infusion. Indirect sources include contamination of animal derived feed with water during processing, storage and transmission.¹⁴ The contamination of feed and crops can also occur from faecal recycling as the drug residues gets excreted in the faeces of treated animals and faecal matter of animals is commonly used fertilizer in fields.¹⁵ The major reasons for occurrence of antibiotic residues in animal sourced foods could be: (i) failure to follow a proper withdrawal period which is specific for each antibiotic drug, (ii) extra label use of antibiotics, (iii) poor record keeping of dairy animals, and (iv) mixing of milk from treated animals with the milk from untreated animals.

Consumption of such products with a higher level of antibiotic residues for a prolonged period may impart serious health risk to consumers. In order to avoid serious public health complications arising from irrational use of antibiotic drugs in producer animals, the Food Safety & Standards Authority of India (FSSAI) have established the Maximum

Residual Limits (MRLs) for antibiotic residues in various food of animal origins (Table 1).¹⁶ Food commodities found to be exceeding these limits are not legally permitted to be sold to consumers in Indian domestic market. Similarly, other food safety agencies across the world such as European commission, US FDA, Codex Alimentarius Commission etc. have also established tolerance limits for antibiotics and other residues in milk to safeguard human health.

To tackle the issue of antibiotic misuse and AMR in India, Government of India and many NGOs have come forward with many initiatives such as an Inter-sectoral coordination committee to regulate non-therapeutic use of antibiotics in food animals has been constituted under National Policy for containment of AMR. Schedule H1 has been implemented by CDSCO and there is blanket ban on over the counter sale of almost 25 antimicrobials including cephalosporin, carbapenems, anti TB drugs & newer fluoroquinolones etc. A road map to tackle global challenge of AMR from Indian perspective was formulated in Chennai Declaration. GARP (Global Antibiotic Resistance Partnership) involving India, Kenya, South Africa, Vietnam, China are among other actions taken in this regard.

Table 1: Tolerance limit of various Antibiotic drug residues in Bovine milk.

Name of Antibiotics	Tolerance limit (mg/kg)
Monensin	0.002
Sulphadimidine	0.025
Ampicillin, Cloxacillin, Salinomycin, Tyvalosin Tartrate, Virginiamycin, Sulphadiazine, Sulphathiazole sodium, Trimethoprim, Sulphadiazine, Sulphanilamide	
Sulphaguanidine, Bacitracin, Apramycin, Cephapirine, Clopidol, Enrofloxacin, Ethopabate, Flavomycin, Sulphaquinoxaline	0.01
Streptomycin	0.02
Colistin	0.05
Ceftiofur	0.1
Lincomycin	0.15
Oxytetracycline/Tetracycline, Spectinomycin	0.1
Neomycin	1.5

Public Health Concern of Antibiotic Residues in Food of Animal Origin

Antibiotics, if consumed for a prolonged period of time even in sub-therapeutic or low doses, may

cause variety of effects on exposed population. Most common direct effects are hypersensitivity reaction (penicillin), yellowing of teeth (tetracycline), aplastic anemia (chloramphenicol) and gastro-intestinal disturbances.¹⁷ Acute toxicity of sulphonamides leads to renal dysfunction and blood dyscrasias, while chronic toxicity leads to conditions like hepatic necrosis, hypothermia. Tetracycline toxicity can also cause hepatotoxicity & nephrotoxicity. Streptomycin causes ototoxicity.¹⁸ Certain macrolides are hepatotoxic in nature.¹⁹ One of the major concerns is transfer of antibiotic resistance to humans which can either be by transfer of antibiotic resistant bacteria (ARB) or Antibiotic resistant gene (ARG) to humans through food chain or livestock handlers.²⁰ AMR is possibly the single biggest threat facing the world in the area of infectious diseases. Although little is known about magnitude of this problem with animal husbandry practices as determinant. This may be due to rare screening offered by the farmers, limited testing by veterinary diagnostic labs, poor surveillance mechanisms in exclusive animal pathogens compared to zoonotic pathogens, lack of resources, cost involved in culture or sensitivity testing, lack of coordination for collection, culture & antimicrobial testing methods or may be due to perceived low priority. However, this serious issue should be viewed at the same level of priorities as climate change or terrorism.

Methods for Detection of Antibiotic Residues in Milk

A plethora of analytical methods have been devised to qualitatively and quantitatively detect occurrence of antibiotic residues in food of animal origin especially milk. The various confirmatory and screening methods can be broadly grouped into four main categories viz. (i) microbiological methods based on bacterial growth inhibition, (ii) Immunochemical techniques (ELISA), (iii) Chromatographic methods (HPLC), and (iv) Biosensors based techniques.

Confirmatory tests rely on primarily on liquid chromatography coupled with diode array detector (HPLC-DAD) or with mass spectrometry (LC-MS).²¹ LC-MS, a type of quantitative assay requires instrument with high response value & high sensitivity (Table 2). LC-MS method can detect multiple analytes in a single run due to its high selectivity & can detect analyte as low as ng/L or ng/gm.²²⁻²³ Various preparation methods such as solvent-phase extraction (SPE) series

like Solid-phase Microextraction (SPME) and Molecularly Imprinted Dispersive solid-phase extraction (MIDSPE) & Solvent Extraction (SE) series like Pressurized liquid extraction (PLE), Accelerated solvent extraction (ASE), and Solid-Liquid extraction (SLE) are implemented before chromatographic analysis by HPLC. Another method which is quick, easy, cheap, effective, rugged & safe (QuEChERS) has been widely used due to its low solvent consumption & high recovery.²⁴ It is a green chemistry based approach wherein, minimum quantity of organic solvents are utilized for extraction of targeted analytes.

Screening methods on the other hand are a type of qualitative or semiquantitative microbiological assay which are based on the reaction between

susceptible organism and targeted antibiotic. Their advantages over the quantitative assay are that these are reliable, cost effective & simple to perform.²⁵ The other semi quantitative immunoassays are ELISA, Fluoroimmunoassay (FIA) & Time resolved Fluoroimmunoassay (TRFIA), Fluorescence-polarization immunoassay (FPIA), Immunosensors & Biosensors.²⁵⁻²⁶ A biosensor, as defined by IUPAC, is a self contained integrated device that is capable of providing specific quantitative or semi quantitative analytical information using a biological recognition element (biochemical receptor), which is retained in direct spatial contact with a transduction element. Biosensors are classified according to the physicochemical transduction elements in electrochemical, optical, mass sensitive and thermal sensors.

Table 2: Detection of antibiotic residues in milk using HPLC.

Compounds	Sample preparation	HPLC system	Recovery	LOD (µg/L)	LOQ (µg/L)	Reference
Oxytetracycline and amoxicillin	SPE	HPLC-DAD	88-98%	1.3-2.0	3.8-6.1	27
58 Antibiotics	LLE	LC-ESI-MS/MS (+)	*	0.06-13.6	*	28
5 Fluoroquinolones	SPE	LC-ESI-MS/MS (+)	*	0.2	0.5	29
23 antibiotics	LLE - SPE	LC-ESI-MS/MS (+)	22 -143%	*	*	30
36 Antibiotics	LLE	TLX-ESI-MS/MS using optimized turbulent flow (+)	60 -105%	0.3-25.0	1.0-75.0	31
6 β-lactam antibiotics	LLE	LC-ESI-MS/MS (+)	42 - 81%	0.4-10.0	1.0-25.0	32
5 Fluoroquinolones and 1 quinolone	LLE	LC-DAD-ESI- MS (+)	64 - 96%	2.4-30	8-110	33
21 antibiotics	LLE	LC-ESI-MS/MS (+)	67 - 128%	0.1-2.5	0.1-5	34
33 Antibiotics	SPE	LC-ESI-MS/MS (+) and (-)	*	0.01-3.70	*	35
38 Antibiotics	LLE - SPE	LC-ESI-MS/MS (+)	68- 118%	0.01-5	0.03-10	36
6 Antibiotics	QuEChERS	LC-ESI-MS/MS (+)	84 -116%	0.4-3	0.1-0.5	37
25 Antibiotics	modified LLE method	LC-ESI-MS/MS (+)	62 -108%	0.2-10	2.5-25	38
6 Cephalosporins	MISPE	LC-ESI-MS/MS (+)	15 - 100%			
± 6%	0.1-3.8	0.4-12.5	39			
61 Antibiotics	LLE-SPE	LC-ESI-MS/MS (+)	62- 119%			
± 10%	0.003-1.57	0.001-5.18	40			
16 Macrolides and metabolites	QuEChERS	LC-ESI-MS/MS (+)	62 - 115%	0.30-0.85	1.1-4.0	41
15 Antibiotics	SPE	CZE-Q-TOF-MS	76 - 106%			
± 13%	0.5-2.9	1.5-9.6	42			
20 Antibiotics	dSPE	LC-ESI-MS/MS (+)	70 - 97%			
± 10%	0.10-2.40	0.33-7.92	43			
4 Antibiotics	LLE	LC-ESI-MS/MS (+)	61 - 111%	0.1-0.5	1.0-5.0	44

* Not available

Conclusion

Since antibiotics play a major role in keeping the livestock sector free from diseases, their occurrence in the foods of animal origin is also a major public health hazard & one of the alarming situations due to rise in number on antimicrobial resistant microorganisms, which often becomes a challenging task to treat. Veterinarians can play a major role in controlling the emergence of antibiotic resistant bacteria by judiciously using antibiotics with broad spectrum action and shorter withdrawal time. Livestock owners should be made aware about the consumption of produce from animal which is under treatment or have been recently treated with antimicrobials. Since, it may not be practically feasible to nullify the use of antibiotics in animals as the pathogens are omnipresent but precautions should always be taken while administering such drugs to animals. The ban on over the counter sale of antibiotics to avoid irrational use by livestock owners, restricting use of drugs in sub-therapeutic doses, avoid mixing of antibiotics in feed as growth promoters are some of the strategies which can probably minimize the risks associated with antibiotic residues in foreseeable future. In advance clinical settings, it is advisable to initiate antibiotic treatment only after the laboratory report of antimicrobial sensitivity testing. Action plans need to be devised in order to control such conditions. We cannot control the evolutionary development of microorganisms which are turning resistant to antibiotics on their own but the least we can do is to avoid the indiscriminate use of antibiotics which is an enabling factor for them to develop resistance against these magic bullets. Newer and old revisited approaches can move us from the end of antibiotic era towards a new dawn of antibacterial agents.

References

- Hattimare D, Shakya S, Patyal A, Chandrakar C, Kumar A. Occurrence and exposure assessment of Aflatoxin M1 in milk and milk products in India. *Journal of Food Science and Technology* 2021. DOI: <https://doi.org/10.1007/s13197-021-05265-4>
- Pandey AK, Shakya S, Patyal A, Ali SL, Bhonsle D, Chandrakar C, Kumar A, Khan R, Hattimare D. Detection of Aflatoxin M1 in bovine milk from different agro-climatic zones of Chhattisgarh, India using HPLC-FLD and assessment of human health risks. *Mycotoxin Research* 2021; 37 (3): 265-73. DOI: <https://doi.org/10.1007/s12550-021-00437-91>
- Kumar A, Nagal KB. Bacterial Contamination of Livestock Feed and its Public Health Significance. *Indian Veterinary Journal* 2014; 91(03): 64-65.
- Kumar A, Gill JPS, Bedi JS, Chhuneja PK. Residues of antibiotics in raw honeys from different apiaries of Northern India and evaluation of human health risks. *Acta Alimentaria, An International Journal of Food Science* 2020; 49(3): 314-20.
- Kumar A, Gill JPS, Bedi JS, Chhuneja PK, Kumar A. Determination of antibiotic residues in Indian honeys and assessment of potential risks to consumers. *Journal of Apicultural Research* 2020; 59(1): 25-34. DOI: 10.1080/00218839.2019.1677000
- Kumar A, Gill JPS, Bedi JS and Kumar A. Pesticide residues in Indian raw honeys, an indicator of environmental pollution. *Environmental Science and Pollution Research* 2018; 25 (34): 34005-34016. DOI: 10.1007/s11356-018-3312-4.
- Kumar A, Gill JPS, Bedi JS. Multiresidue determination of pesticides in market honey from northern India using QuEChERS approach and assessment of potential risks to consumers. *Current Science* 2018; 115(2): 283-91.
- Gupta A, Nagal KB, Panda AK. Bacteriological quality and safety of raw milk and milk products with special reference to coliforms in Palampur (Himachal Pradesh). *Himachal Veterinary Journal* 2013; 6 (1): 98-101.
- Panda AK, Kumar A, Thakur SD, Shalmali. Evaluation of bacteriological quality of meat and meat products in Palam Valley of North Western Himalayas. *Journal of Veterinary Public Health* 2012; 10 (1): 21-25.
- Phillips I, Casewell M, Cox T, De Groot B, Friis C, Jones R, Nightingale C, Preston R, Waddell J. Does the use of antibiotics in food animals pose a risk to human health? A critical review of published data. *Journal of Antimicrobial Chemotherapy* 2004;53:28-52.
- Vishnuraj MR, Kandeepan G, Rao KH, Chand, Kumbhar S. Occurrence, public health hazards and detection methods of antibiotic residues in foods of animal origin: A comprehensive review. *Cogent Food & Agriculture* 2016;2:1. DOI: 10.1080/23311932.2016.1235458
- Nisha AR. Antibiotic residues-A global health hazard. *Veterinary World* 2008;1:375-77.
- Kirchhelle C. Pharming animals: A global history of antibiotics in food production (1935-2017). *Nature* 2018;4(1):1-13.
- Kumar A, Patyal A, Panda AK. Sub-therapeutic Use of Antibiotics in Animal Feed and their Potential Impact on Environmental and Human Health: A Comprehensive Review. *Journal of Animal Feed Science and Technology* 2018;6(1):15-25. DOI: <http://dx.doi.org/10.21088/jafst.2321.1628.6118.3>
- Jindal P, Bedi J, Singh R, Gill JP. Epidemiological assessment of antibiotic residues in dairy farm milk and farm waste and water in northern India. *Environmental Science and Pollution Research* 2021;28:29455-66. DOI: <https://doi.org/10.1007/>

- s11356-020-12057-4
16. FSSAI. Food Safety and Standards Authority of India Food Safety and Standards (Contaminants, toxins and residues) Regulations 2011. F. No. 2-15015/30/2010. <https://archive.fssai.gov.in>. Accessed 15 August 2020
 17. Davies J, Davies D. Origins and evolution of antibiotic resistance. *Microbiology and Molecular Biology Reviews* 2010;74:417–33.
 18. Katz SE, Brady MS. Antibiotic residues in food and their significance. *Food Biotechnology* 2000;14:147–71.
 19. Dewdney J, Maes L, Raynaud J, Blanc F, Scheid J, Jackson T, Lens S, Verschueren C. Risk assessment of antibiotic residues of β -lactams and macrolides in food products with regard to their immuno-allergic potential. *Food and Chemical Toxicology* 1991;29:477–83.
 20. Levy SB, Marshall B. Antibacterial resistance worldwide: causes, challenges and responses. *Nature Medicine* 2004;10:S122.
 21. Blasco C, Di Corcia A, Picó Y. Determination of tetracyclines in multi-specie animal tissues by pressurized liquid extraction and liquid chromatography-tandem mass spectrometry. *Food Chemistry* 2009;116:1005–12.
 22. Stolker A, Brinkman UT. Analytical strategies for residue analysis of veterinary drugs and growth-promoting agents in food producing animals-a review. *Journal of Chromatography A* 2005;1067(1–2):15–53.
 23. Kittlaus S, Schimanke J, Kempe G, Speer K. Development and validation of an efficient automated method for the analysis of 300 pesticides in foods using two-dimensional liquid chromatography tandem mass spectrometry. *Journal of Chromatography A* 2013;1283:98–109.
 24. Ridgway K, Lalljie SP, Smith RM. Sample preparation techniques for the determination of trace residues and contaminants in foods. *Journal of Chromatography A* 2007;1153 (1–2):36–53.
 25. Cháfer-Pericás C, Maquieira A, Puchades R. Fast screening methods to detect antibiotic residues in food samples. *Trends in Analytical Chemistry* 2010;29:1038–49.
 26. Kantiani L, Farré M, Barceló D, Barceló D. Analytical methodologies for the detection of β -lactam antibiotics in milk and feed samples. *Trends in Analytical Chemistry* 2009;28(6):731.
 27. Kumar A, Panda AK, Sharma N. Determination of antibiotic residues in bovine milk by HPLC-DAD and assessment of human health risks in North-western Himalayan region, India. *Journal of Food Science & Technology* 2021. DOI: <https://doi.org/10.1007/s13197-021-04988-8>
 28. Gaugain-Juhel M, Delepine B, Gautier S, Fourmond MP, Gaudin V, Hurtaud-Pessel D, Verdon E, Sanders P. Validation of a liquid chromatography-tandem mass spectrometry screening method to monitor 58 antibiotics in milk: A qualitative approach. *Food Additives & Contaminants: Part A* 2009;26(11):1459–71.
 29. Tang Q, Yang T, Tan X, Luo J. Simultaneous determination of fluoroquinolone antibiotic residues in milk sample by solid-phase extraction-liquid chromatography-tandem mass spectrometry. *Journal of Agricultural and Food Chemistry* 2009;57(11):4535–39.
 30. Clark SB, Storey JM, Turnipseed SB. Optimization and validation of a multiclass screening and confirmation method for drug residues in milk using high-performance liquid chromatography/tandem mass spectrometry. *Journal of AOAC International* 2011;94(2):383–93.
 31. Bousova K, Senyuva H, Mittendorf K. Multiresidue automated turbulent flow online LC-MS/MS method for the determination of antibiotics in milk. *Food Additives & Contaminants: Part A* 2012;29(12):1901–12.
 32. Jank L, Hoff R, Tarouco P, Barreto F, Pizzolato T. β -lactam antibiotics residues analysis in bovine milk by LC-ESI-MS/MS: A simple and fast liquid-liquid extraction method. *Food Additives & Contaminants: Part A* 2012;29(4):497–507.
 33. Ruiz-Viceo J, Rosales-Conrado N, Guillén-Casla V, Pérez-Arribas L, León-González M, Polo-Díez L. Fluoroquinolone antibiotic determination in bovine milk using capillary liquid chromatography with diode array and mass spectrometry detection. *Journal of Food Composition and Analysis* 2012;28(2):99–106.
 34. Tang YY, Lu HF, Lin HY, Hin YC, Hwang DF. Development of a quantitative multi-class method for 18 antibiotics in chicken, pig, and fish muscle using UPLC-MS/MS. *Food Analytical Methods* 2012;5(6):1459–68.
 35. Freitas A, Barbosa J, Ramos F. Development and validation of a multi-residue and multiclass ultra-high-pressure liquid chromatography-tandem mass spectrometry screening of antibiotics in milk. *International Dairy Journal* 2013;33(1):38–43.
 36. Han R, Zheng N, Yu Z, Wang J, Xu X, Qu X, Li S, Zhang Y, Wang J. Simultaneous determination of 38 veterinary antibiotic residues in raw milk by UPLC-MS/MS. *Food Chemistry* 2015;181:119–26.
 37. Wang YL, Liu ZM, Ren J, Guo BH. Development of a method for the analysis of multiclass antibiotic residues in milk using QuEChERS and liquid chromatography-tandem mass spectrometry. *Foodborne Pathogens and Disease* 2015; 12(8):693–703.

38. Martins MT, Barreto F, Hoff RB, Jank L, Arsand JB, Motta TMC, Schapoval EES. Multiclass and multi-residue determination of antibiotics in bovine milk by liquid chromatography tandem mass spectrometry: Combining efficiency of milk control and simplicity of routine analysis. *International Dairy Journal* 2016;59:44–51.
39. Baeza AN, Urraca JL, Chamorro R, Orellana G, Castellari M, Moreno-Bondi MC. Multiresidue analysis of cephalosporin antibiotics in bovine milk based on molecularly imprinted polymer extraction followed by liquid chromatography tandem mass spectrometry. *Journal of Chromatography A* 2016;1474:121–29.
40. Tian H, Wang JQ, Zhang YD, Li SL, Jiang JD, Tao DL, Zheng N. Quantitative multiresidue analysis of antibiotics in milk and milk powder by ultra-performance liquid chromatography coupled to tandem quadrupole mass spectrometry. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences* 2016;1033:172–79.
41. Zhou W, Ling Y, Liu T, Zhang Y, Li JZ, Li HN, Wu WJ, Jiang SJ, Feng F, Yuan F, Zhang F. Simultaneous determination of 16 macrolide antibiotics and 4 metabolites in milk by using Quick, Easy, Cheap, Effective, Rugged, and Safe extraction (QuEChERS) and high-performance liquid chromatography tandem mass spectrometry. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences* 2017;1061:411–20.
42. Moreno-Gonzalez D, Hamed AM, Gilbert-Lopez B, Gamiz-Gracia L, Garcia-Campana AM. Evaluation of a multiresidue capillary electrophoresis-quadrupole-time-of-flight mass spectrometry method for the determination of antibiotics in milk samples. *Journal of Chromatography A* 2017;1510:100–7.
43. Wang L, Yang BX, Zhang XS, Zheng HG. Novel two-dimensional liquid chromatography-tandem mass spectrometry for the analysis of twenty antibiotics residues in dairy products. *Food Analytical Methods* 2017;10(6):2001–10.
44. Magon T, da Silveira R, Galuch MB, Fagan EP, Feitoza AFD, Palombini SV, Santos OO, Visentainer JV. Simultaneous determination of four antibiotics in raw milk by UPLCMS/ MS using protein precipitation as sample preparation: Development, validation, and application in real samples. *Journal of the Brazilian Chemical Society* 2018;29(11):2441–8

