

REVIEW ARTICLE

Forensic Application of Non-destructive ATR-FTIR Spectroscopic Technique for Organophosphorus Pesticides Analysis

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ABSTRACT

Over a long period organophosphorus pesticides (OPPs) such as Tusk, Dimex, Dysac, Sacban and Monovip contain Malathion, Dimethoate, Dichlorovos, Chlorpyrifos and Monochrotophos as active chemicals, respectively are used in agriculture to enhance the crops yield. However, these chemicals are extensively used in criminal cases such as suicidal, accidental, and homicidal throughout the world. Such cases can conveniently be resolved with the help of spectral library as reference/standard to compare with unknown or suspected samples recovered from scene of crime. The spectral library was generated using ATR-FTIR technique which reliable and non destructive in nature. This spectral library has been applied to identify the OPPs in different biological samples and investigate, characterize the OPPs. ATR-FTIR spectroscopic data are, therefore, a prerequisite for their identification. Finally, we believe that the use of ATR-FTIR should make it possible to identify all tested OPPs in one single analysis, even in the low ppm concentration range.

KEYWORDS | ATR-FTIR, organophosphorus, biological samples
non-destructive, spectral library

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INTRODUCTION

PESTICIDES PROTECT THE CROPS from pests but their usage is harmful to humans and other animals due to potential toxicity. Only 1% of the total amount of pesticides (approx. 4 millions tones) annually applied reaches to target pests.¹ On the basis of targets, pesticides are divided into several types such as herbicides, insecticides, fungicides, rodenticides and others. Over long period, organophosphorus pesticides (OPPs) or organophosphates are being used for protecting the crops and also as warfare agents. Organophosphorus pesticides have a pentavalent phosphorous atoms attached to a sulfur or oxygen atoms by double covalent bond predicted by a number of chemicals e.g. Malathion, Dimethoate, Dichlorovos, Chlorpyrifos, Monochrotophos etc.

(Fig. 1). OPPs are named on the basis of type of group attached to phosphorous such as phosphate, thiono, thiol and thithiol type. R1 and R2 predict different substituent groups (alkoxy, alkyl, amino, thioalkyl etc.) present at the skeletal base of OPPs.^{2,3} R1 and R2 are replaced by propyl and ethyl groups to manage resistant pests respectively. X represents the alkyl, alkoxy, aryl, heterocyclic, aryl oxy, arylthio and alkylthio groups. OPPs are structurally similar within a class and have a phosphorous atoms with characteristics phosphoryl bond (P=O) OR thiophosphoryl bond (P=S).^{1,4} OPPs and their metabolites are used as a pesticide across the world population. The WHO (World Health Organization) was reported that every year 3 million peoples were affected



How to cite this article

Shivpoojan Kori. Forensic Application of Non-destructive ATR-FTIR Spectroscopic Technique for Organophosphorus Pesticides Analysis. Indian J Forensic Med and Pathol. 2021;14 (2Special):395-402

by the acute poisoning of OPPs. OPPs are also caused neurotoxicity due to inhibition of acetyl cholinesterase (AChE) enzymes and butyryl cholinesterase (BuChE) enzymes, present in the synaptic membrane of CNS (central nervous system) and PNS (peripheral nervous system) of vertebrates. AChE and BuChE enzymes hydrolyze the neurotransmitter acetylcholine into acetyl and choline which regulates the signal transmission by sodium channel in the CNS and PNS system. OPPs block the active sites of AChE and BuChE and finally deactivate the acetylcholine (ACh), leads to inhibition cholinergic neurotransmission due to accumulation of the acetylcholine. These neurotoxic effects results in neuromuscular paralysis.^{5,6} OPPs are also potent to causes immunotoxicity, genotoxicity and carcinogenicity, finally leading to death. Conclusively, OPPs are the most dangerous and determined organic pollutants chemicals due to their potential toxicity. Recently, most of the criminal cases are taking place due to pesticide toxicity. In forensic science set-ups, these types of cases are analyzed from blood, foodstuff, spits and the chemical bottles

pesticides poisoning have been reported in rural area of Asia.¹⁰ WHO (1990) had published the data and found 3 millions/year detectable cases related to pesticides poisoning resulting in 220,000 deaths^{11,12} OPPs categories of pesticides had been screened mostly in forensic science laboratory using thin layer chromatography (TLC). Criminal cases related to pesticides are occurring daily and are being forwarded to forensic science laboratory for pesticides analysis using thin layer chromatography and are further confirmed by using FT-IR and GC-MS techniques. GC-MS is destructive, costly and time consuming technique so cannot be used routinely in laboratory. However, FT-IR is non-destructive and reliable technique. Therefore, it has been planned to analyze the ATR-FTIR spectrum of commonly available OPPs, so as to obtain the standard or reference data and this spectral library has been applied in real spiked biological samples. The studies have been carried out using ATR-FTIR spectroscopic technique. It is a powerful technique for analysis of trace and bulk constituents of matrices sample. Infrared absorption technique has efficiently been

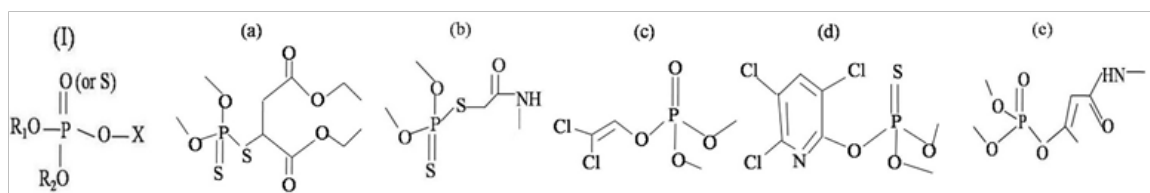


Figure 1: (i) General chemical structure of OPPs and (a) Malathion (b) Dimethoate (c) Dichlorovos (d) Chlorpyrifos (e) Monochrotophos.

Source: Author self.

seized from crime scene to know the cause. Few OPPs such as Malathion, Dimethoate, dichlorovos, Chlorpyrifos and Monochrotophos are highly toxic in nature and have been used in criminal cases.⁷ In Brazilian institute, research groups have been analyzed the acetyl cholinesterase enzymes inhibitors such as aldicarb, OPPs chemicals and confirming using FT-IR technique for forensic purpose.⁸ In pesticide poisoning cases, OPPs are the most detectable in forensic intoxication viz. 63% and out of it, Quinalphos was the most common OPPs responsible for 28.8% of the total positive cases.⁹ Suicidal cases are most common and approximate 60% cases of death due to

used to detect the microscopic pesticide residue.¹³ The technique is non-destructive which makes it, the best suitable for forensic utility because, the re-analysis of sample can be carried for future purpose.

METHOD

Five OPPs—Malathion, Dimethoate, Dichlorovos, Chlorpyrifos and Monochrotophos—were analyzed and characterized using ATR-FTIR spectroscopic technique.

Sample Preparation

Human biological control samples (free from milk, saliva and vomit materials) were obtained

from healthy volunteer and kept frozen at -20°C . 500 μL volume of each biological samples were spiked three times ($n=3$) with each OPPs (Malathion, Dimethoate, Dichlorovos, Chlorpyrifos and Monochrotophos) up to 0.5ppm concentration in a scaled centrifugal vial. Samples were vortexed for FTIR analysis. Each sample was divided into four groups. First three groups were spiked with pesticides at the same concentration (0.5 ppm). However, pesticides were not spiked in the last group and served as a control.

Standard stock solution (2000ppm) was prepared for each OPPs separately. Tusk (50% EC), Dimex (30% EC), Dysac (76% EC), Sacban (20% EC) and Monovip (36% EC) contain Malathion, Dimethoate, Dichlorovos, Chlorpyrifos and Monochrotophos as active chemicals, respectively. OPPs pesticides were purchased from Shivalik agrochemicals at Chandigarh (India).

ATR-FTIR Spectroscopy

For ATR-FTIR spectra of each sample, four scans were recorded with a resolution of 4 cm^{-1} , in the range from 4,000 to 700 cm^{-1} using ATR-FTIR, (the Agilent Cary 630 FTIR Spectrometer). The spectral data was analyzed using Agilent Microlab FTIR software. 2.0 μL samples were placed directly on the internal reflecting diamond crystal of FTIR with the help of micropipette.

RESULTS AND DISCUSSIONS

A number of methods had been used to analyse the organophosphorous pesticides (OPPs) residue in suspected samples of forensic interest. Use of infrared spectroscopy in forensic science is best technique because of sample is analysed without destruction & treatment. In spite of these efforts, an attempt in the present study has been equipped to obtain chemical fingerprinting of some commonly encountered OPPs using a non-destructive ATR-FTIR method. The main motive is to generate ATR-FTIR spectra database of these OPPs due to makes it possible to positively identify the major OPPs found at crime scene. Finally, we believe that the use of ATR-FTIR should make it possible to identify all tested OPPs in one single analysis, even in the low ppm concentration range.

This study was to propose the spectral library of OPPs as a reference for any forensic chemist and toxicologist in field of forensic science.

The generated ATR-FTIR spectral library at wavenumber (cm^{-1}) was depicted in Table 1 and ATR-FTIR spectrum of spiked OPPs in biological samples was depicted in Table 3. The vibrational assignment of the signature peaks of each OPPs has been discussed in table 2. IUPAC name, Chemical structure and molecular formula of These reference OPPs compounds have been described which are used in this study such as (i) Tusk: The active component of Tusk is Malathion ($\text{C}_{10}\text{H}_{19}\text{O}_6\text{P}_2\text{S}_2$) with IUPAC name Diethyl 2-[(dimethoxyphosphorothioyl) sulfanyl] butanedioate. Chemical structure was depicted in Fig. 1(a). ATR-FTIR spectrum was shown in Fig. 2(a). The vibrational assignment of the principle peaks of ATR-FTIR spectrum has been depicted in Table 2.¹⁴ (ii) Dimex: It contains Dimethoate ($\text{C}_2\text{H}_{12}\text{NO}_3\text{P}_2\text{S}_2$) which chemically is designated as O, O-dimethyl S-[2-(methyl amino)-2-oxoethyl] dithiophosphate. Structurally, Dimethoate was depicted in Fig. 1(b) and ATR-FTIR spectrum was shown in Fig. 2 (b). (iii) Dysac: Active component in Dysac is Dichlorovos ($\text{C}_4\text{H}_7\text{Cl}_2\text{O}_4\text{P}$) with specific chemical name as 2, 2-dichlorovinyl dimethyl phosphate. The chemical structure of Dysac was shown in Fig. 1(c). Fig. 2 (c) depicts the FT-IR spectrum.¹⁴⁻¹⁶ (iv) Sacban: The commercial product contains Chlorpyrifos ($\text{C}_9\text{H}_{11}\text{Cl}_3\text{NO}_3\text{PS}$). It is chemically labeled as O,O-Dethyl O-3,5,6-trichloropyridin-2-ylphosphorothioate [Fig. 1(d)]. The FT-IR spectrum was shown in Fig. 2 (d). Table 2 shows the vibrational assignment of chlorpyrifos.^{13,15,17,18} (v) Monovip: It contains Monochrotophos ($\text{C}_7\text{H}_{14}\text{NO}_5\text{P}$) which chemically is represented as Dimethyl (E)-1-methyl-2-(methylcarbomyl) vinyl phosphate (Fig. 1 (e)). FT-IR spectrum of product was shown in Fig. 2 (e). Table 2 shows the vibrational assignment of monochrotophos.^{12,14,15,19,20} This OPPs spectral library has been employed to match the peaks of OPPs residue in spiked biological samples.

In this work, ATR-FTIR spectroscopy has been used to generate a spectral library of some organophosphorous pesticides of forensic interest. These types of pesticides can be seized as bulk market pesticides as well as traces found at the crime scene. There can be legal queries to identify OPPs to forensic expert has to answer. In these

cases, the standard reference database is required for comparison. The modern FTIR systems with diamond ATR proves to be a rapid, sensitive and non-destructive analysis of samples with very little effort. This spectral library can be used as a reference library when an unknown sample is suspected of being organophosphorous. Some of the closely related structures may have spectra very similar to that of particular pesticides, especially for organophosphorous. Thus, it is imperative that appropriate techniques should be used by the forensic chemist and toxicologist to ensure the optimal spectral data are obtained to get a feel for the discriminating features of a spectrum. Screening of pesticides using ATR-FTIR technique is particularly convenient because of both its speed and ability to use without sample preparation.

CONCLUSIONS

FT-IR is a very sensitive, rapid, easy, reliable, and non-destructive technique. Application of non-destructive analytical technique to analyze the forensic evidence as samples of forensic interest without destroying the sample. ATR-FTIR is a beneficial to forensic scientist/expert in field of forensic science. This generated spectral library of OPPs by ATR-FTIR can be used as reference/standard for comparison of pesticides products, seized from crime scene.

In this work, ATR-FTIR spectral library was generated for some OPPs of forensic interests which had been frequently used in criminal cases and mostly recovered from crime scene. This

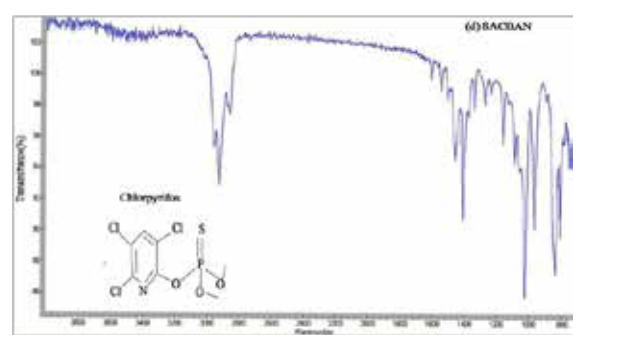
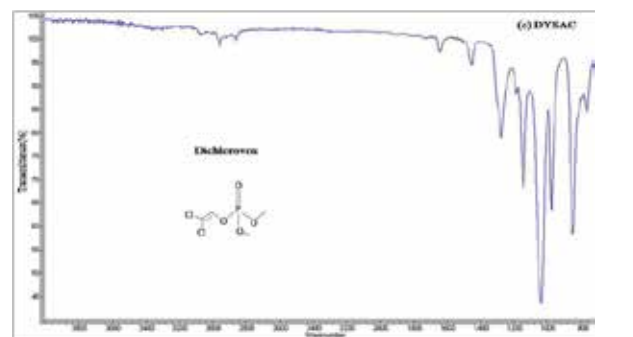
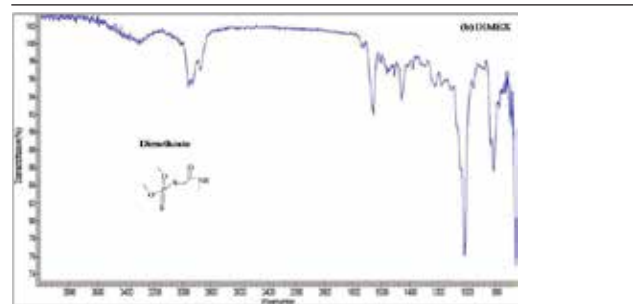
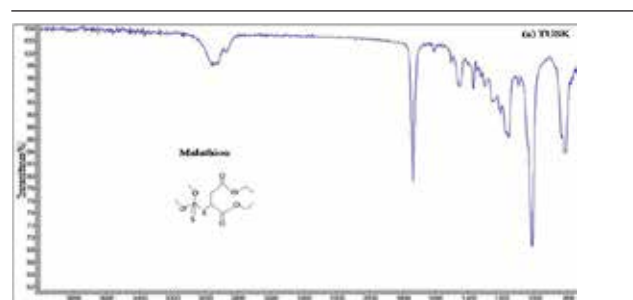
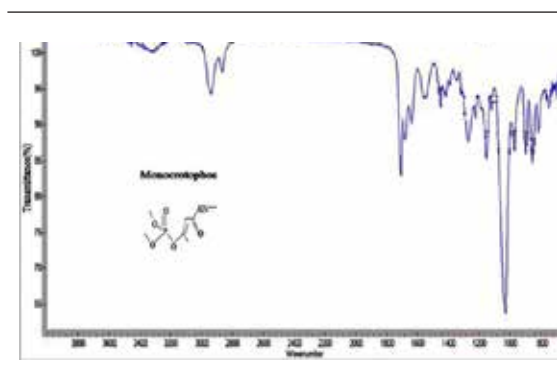


Figure 2(a): FTIR spectrum of Tusk OPPs product.

Figure 2(b): FTIR spectrum of Dimex OPPs product.

Figure 2(c): FTIR spectrum of Dysac OPPs product.

Figure 2(d): FTIR spectrum of Sacban OPPs product.

Figure 2(e): FTIR spectrum of Monocrotophos OPPs product.

Figure 2(f): FTIR spectrum of Monocrotophos OPPs product.

Source: Author Self

| PURE OPPS | WAVENUMBERS (CM-1) |
|---------------------------------|---|
| Tusk 20% EC (Malathion) | 2869(w), 1735(s), 1456(m), 1372(w), 1255(w), 1157(m), 1014(s), 817(m) |
| Dimex 30% EC (Dimethoate) | 2948(m), 2874(m), 1661(s), 1550(w), 1457(m), 1296(w), 1221(w), 1176(w), 1016(s), 810(m) |
| Dysac 76% EC (Dichlorovos) | 1644 (w), 1456(w), 1279(m), 1147(m), 1039, 977(m), 857(m), 765(w) |
| Sacban 20% EC (Chlorpyrifos) | 2924(s), 2857(w), 1607(w), 1544(w), 1506(w), 1457(w), 1410(s), 1338(w), 1214(w), 1162(w), 1024(s), 984(m), 835(s), 805(s) |
| Monovip 36% EC (Monochrotophos) | 2937(w), 2862(w), 1706(s), 1883(w), 1637(w), 1550(w), 1449(w), 1270(m), 1154(m), 1033(s), 970(w), 898(w), 855(w), 811(w) |

Table 1: ATR-FTIR spectral peaks of OPPs of standard at transmittance (%) vs. wavenumber (cm-1) in OPPs products.

| PURE OPPS | WAVENUMBERS (CM-1) | ASSIGNMENT |
|------------------------------|---------------------------------|---|
| Tusk 20% EC (Malathion) | 2869cm-1 | aliphatic C-H stretching vibration |
| | 1735cm-1 | (C=O) carbonyl stretching vibration of ester group |
| | 1456cm-1 | C-H stretching vibrations of CH ₂ |
| | 1370cm-1 | C-H Vibration of CH ₃ group |
| | 1255cm-1 | C-H rocking vibration of CH ₂ |
| | 1157cm-1 | C-O vibration |
| | 1014cm-1 | P-O stretching of P-O-CH ₃ group |
| | 817cm-1 | P-O vibration of additional phosphates (P043-) group |
| Dimex 30% EC (Dimethoate) | 2948cm-1 | aliphatic C-H asymmetric stretching vibration of CH ₃ |
| | 2874cm-1 | aliphatic C-H symmetric stretching vibration of CH ₃ |
| | 1661cm-1 | (C=O) carbonyl stretching vibration |
| | 1550cm-1 | P=S bending vibration |
| | 1457cm-1 | C-H stretching vibrations of CH ₂ |
| | 1310cm-1 | C-H Vibration of CH ₃ group |
| | 810cm-1 | P-O stretching of P-O-CH ₃ group, P-O vibration of additional phosphates (P043-) group bands |
| | 1296cm-1, 1221cm-1, 1176cm-1 | C-N stretching vibration |
| Dysac 76% EC (Dichlorovos) | 1644cm-1 | C=C stretching in alkenes |
| | 1456cm-1 | C-H stretching vibrations of CH ₂ |
| | 1039cm-1 | P-O stretching of P-O-CH ₃ group |
| | 977cm-1 | C-H bending vibration of CH ₂ |
| | 857cm-1 | P-O vibration of additional phosphates (P043-) group |
| | 765 cm-1 | C-Cl scissoring vibration |
| | 1279cm-1, 1147cm-1 | C-O bending vibration |
| Sacban 20% EC (Chlorpyrifos) | 1607 to 984cm-1 | C=N stretching, pyridine stretching, ring vibration, ring breathing, Cl-C stretching, trigonal ring breathing and P=S stretching |
| | 2924cm-1 | C-H asymmetric stretching vibration of CH ₃ |
| | 2857cm-1 | C-H symmetric stretching vibration of CH ₃ |
| | 1607cm-1 | C=C Vibration |
| | 1544cm-1 | P=S stretching |
| | 1410cm-1 | CH ₂ bending |
| | 1338cm-1 | C-O stretching |
| | 1024cm-1 | CH ₂ in plane vibration |
| | 984cm-1 | CH ₃ wagging vibration, P-O stretching of P-O-CH ₃ and P-O vibration of additional phosphates (P043-) group bands |
| | 1506cm-1, 1457cm-1 | vibration mode of CH ₂ and CH ₃ group |
| | 1214cm-1, 1162cm-1 | C-N stretching vibration |
| | 835cm-1, 805cm-1 | C-Cl rocking vibration |
| | Monovip 36% EC (Monochrotophos) | 2937cm-1 |
| 2862cm-1 | | C-H symmetric stretching vibration of CH ₂ |
| 1706cm-1 | | (C=O) carbonyl stretching vibration |
| 1683cm-1 | | peaks of additional (C=O) carbonyl stretching vibration |
| 1637cm-1 | | C=C stretching in alkenes |
| 1550cm-1 | | N-H bending vibration |
| 1449cm-1 | | bending vibration of CH ₂ |
| 1033cm-1 | | P-O stretching of P-O-CH ₃ |
| 970cm-1 | | N-CO bending at out of plane |
| 898cm-1 | | out of plane bending CH in C-CN |
| 1270cm-1, 1154cm-1 | | C-N bending |
| 855cm-1, 811cm-1 | | mode of C-CH bending |

Table 2: Vibrational assignment of ATR-FTIR spectral peaks corresponding wavenumber (cm-1) of OPPs.

| SPIKED OPPS | WAVENUMBERS (CM-1) |
|---------------------------------|--|
| Tusk 20% EC (Malathion) | 2869(w), 1735(s), 1372(w), 1157(m), 1014(s), |
| Dimex 30% EC (Dimethoate) | 2948(m), 1661(s), 1550(w), 1457(m), 1176(w), 1016(s), 810(m) |
| Dysac 76% EC (Dichlorovos) | 1644 (w), 1279(m), 1039, 977(m), 857(m), |
| Sacban 20% EC (Chlorpyrifos) | 2924(s), 1607(w), 1506(w), 1410(s), 1338(w), 1214(w), 984(m), 835(s), |
| Monovip 36% EC (Monochrotophos) | 2937(w), 2862(w), 1883(w), 1637(w), 1449(w), 1154(m), 1033(s), 970(w), 855(w), |

(A) ATR-FTIR spectral peaks of OPPs of spiked blood sample

| SPIKED OPPS | WAVENUMBERS (CM-1) |
|---------------------------------|--|
| Tusk 20% EC (Malathion) | 2869(w), 1735(s), 1456(m), 1372(w), 1255(w), 1014(s), 817(m) |
| Dimex 30% EC (Dimethoate) | 2874(m), 1661(s), 1550(w), 1457(m), 1221(w), 1176(w), 1016(s), |
| Dysac 76% EC (Dichlorovos) | 1644 (w), 1279(m), 1147(m), 1039, 977(m), 857(m), 765(w) |
| Sacban 20% EC (Chlorpyrifos) | 2924(s), 1544(w), 1506(w), 1457(w), 1338(w), 1214(w), 1162(w), 1024(s), 835(s), 805(s) |
| Monovip 36% EC (Monochrotophos) | 2937(s), 2862(w), 1706(s), 1637(w), 1550(w), 1270(m), 1033(s), 970(w), 811(m) |

(B) ATR-FTIR spectral peaks of OPPs of spiked urine sample

| SPIKED OPPS | WAVENUMBERS (CM-1) |
|---------------------------------|---|
| Tusk 20% EC (Malathion) | 2869(s), 1735(s), 1372(w), 1157(m), 817(m) |
| Dimex 30% EC (Dimethoate) | 2948(m), 2874(m), 1550(w), 1457(m), 1296(w), 1176(w), 1016(s), |
| Dysac 76% EC (Dichlorovos) | 1644 (s), 1279(m), 1147(m), 1039, 977(m), 857(m), 765(w) |
| Sacban 20% EC (Chlorpyrifos) | 2924(s), 2857(w), 1544(w), 1457(w), 1410(s), 1214(w), 1162(w), 984(m), 805(s) |
| Monovip 36% EC (Monochrotophos) | 2937(w), 2862(w), 1706(s), 1637(w), 1550(w), 1270(m), 1033(s), 970(w), 855(w), 811(w) |

(C) ATR-FTIR spectral peaks of OPPs of spiked milk sample

| SPIKED OPPS | WAVENUMBERS (CM-1) |
|---------------------------------|---|
| Tusk 20% EC (Malathion) | 2869(s), 1735(s), 1372(w), 1157(m), 817(m) |
| Dimex 30% EC (Dimethoate) | 2948(m), 2874(m), 1550(w), 1457(m), 1296(w), 1176(w), 1016(s), |
| Dysac 76% EC (Dichlorovos) | 1644 (s), 1279(m), 1147(m), 1039, 977(m), 857(m), 765(w) |
| Sacban 20% EC (Chlorpyrifos) | 2924(s), 2857(w), 1544(w), 1457(w), 1410(s), 1214(w), 1162(w), 984(m), 805(s) |
| Monovip 36% EC (Monochrotophos) | 2937(w), 2862(w), 1706(s), 1637(w), 1550(w), 1270(m), 1033(s), 970(w), 855(w), 811(w) |

(D) ATR-FTIR spectral peaks of OPPs of spiked saliva sample

| SPIKED OPPS | WAVENUMBERS (CM-1) |
|---------------------------------|---|
| Tusk 20% EC (Malathion) | 2869(w), 1735(s), 1456(m), 1255(w), 1157(m), 1014(s), |
| Dimex 30% EC (Dimethoate) | 2948(m), 1550(w), 1457(m), 1296(w), 1221(w), 1016(s), 810(m) |
| Dysac 76% EC (Dichlorovos) | 1644 (m), 1279(m), 1147(m), 1039, 977(m), 857(m), 765(w) |
| Sacban 20% EC (Chlorpyrifos) | 2924(s), 2857(w), 1544(w), 1506(w), 1457(w), 1410(s), 1338(w), 1162(w), 1024(s), 984(m), 805(s) |
| Monovip 36% EC (Monochrotophos) | 2937(w), 1706(s), 1883(s), 1637(w), 1270(m), 1154(m), 1033(s), 855(w), 811(w) |

(E) ATR-FTIR spectral peaks of OPPs of spiked vomit sample at transmittance (%) vs. wavenumber (cm-1) in OPs products.

(s): strong peak; (w): weak peak; (m): medium peak

generated ATR-FTIR spectral library has been employed in OPPs spiked biological samples. This ATR-FTIR spectral library could be of great help to forensic scientist to characterize and screen the suspect OPPs for data assessment. It is a practically potential to screen out residual OPPs using ATR-FTIR directly. The present work can be scaled up to grant potential positive contribution to discriminate mysterious OPP. **IJFMP**

Recommendation

Present work leaves an off-shoot where, we recommend further, to generate ATR-FTIR spectral library of others standard pesticides and drugs as well as OPPs residue in real samples of forensic interest.

Abbreviations

OPPs: Organophosphorous pesticides, ATR-FTIR: Attenuated total reflection-fourier transforms infra red spectroscopy.

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