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Role of Autologous Platelet Rich Plasma in Tangential Excision and Skin Grafting in Pediatric Scald Burns

Chinthi Reddy Pranaypal¹, Ravi Kumar Chittoria², Neljo Thomas³

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Abstract

Autologous plasma rich plasma (APRP) is an increasingly popular adjunct in surgical, medical and aesthetic interventions. Their beneficial effects lie in their ability to deliver a high concentrate of growth factors. In our study APRP was utilised in a child with scald burns to evaluate the efficacy and mechanism of action of APRP in successful take of a split skin graft.

Keyword: Autologous plasma, Rich plasma, Scaldburns, Grafting

INTRODUCTION

Autologous Platelet Rich Plasma is a blood product rich in platelets, growth factors and chemokines. Since 1990s role of PRP is being discussed as an agent in tissue repair.¹ Now a days it is widely studied for its role in wound healing.² Split skin thickness grafts (STSG) are resurfacing procedures in plastic surgery and it mainly depends on wound bed preparation. Skin graft take involves plasma imbibition, inoculation

and neovascularization. A successful take of graft depends on wound bed vascular status, micro environment and adequate hemostasis. APRP being rich in platelets augments healing process by platelet plug formation, conversion of fibrinogen to fibrin which in turn helps in adhesion of graft and provides a stable fixation. The growth factors from APRP promote angiogenesis and collagen deposition augmenting take of graft. Here in this study, we are evaluating the effect of APRP in successful take of a STSG in a child with scald burns.

MATERIALS AND METHODS

This study was conducted in tertiary care centre in department of plastic surgery after getting the department ethical committee approval. Informed consent was obtained for examination and clinical photography. One year old child presented with accidental scald burns over right upper limb and trunk (Fig. 1). Child was treated with tangential excision (Fig. 2) and split thickness skin graft.

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Wound bed for the graft was prepared by injecting APRP into the wound (Fig. 3). The split skin graft was taken from right thigh of child and grafted into the scald burn (Fig. 4).

Autologous platelet rich plasma (APRP) obtained by standard double centrifugation protocol using

10cc of the patient's blood. Blood was separated into 3 layers namely Platelet poor plasma (PPP) at top, PRP in middle and RBC at bottom. RBC and PPP are discarded sequentially. PRP obtained was added with thrombin. The obtained APRP was injected into scald burns after tangential excision (Fig. 3).



Fig. 1: Child with scald burns



Fig. 2: Tangential excision being done



Fig. 3: Autologous Platelet Rich Plasma being injected into wound



Fig. 4: Split thickness skin graft being applied

RESULTS

After 14 days of tangential excision and skin grafting with APRP, there was good take of graft without any local adverse effects (Fig. 5)



Fig. 5: Postoperative day 14 of skin grafting

DISCUSSION

Autologous platelet rich plasma (APRP) as the name implies refers to the plasma derived from the patient's own blood with a platelet count higher than the platelet count in the peripheral blood of the patient. Historically having been used to treat thrombocytopenia, the use in other specialties became widespread with its use in sports medicine to treat musculoskeletal injuries³. Its use in wound management results from the observation that wounds have a pro inflammatory environment that impairs healing. In addition, wounds have a high protease activity that impairs functioning of growth

factors. APRP used in a chronic wound serves as a source of growth factors and hence has mitogenic, angiogenic and chemotactic properties. APRP has also been shown to stimulate human dermal fibroblast proliferation and thus increasing the deposition of TYPE I collagen, the above mechanism being proposed to its use in scar management. Application of activated APRP also provides 5 to 10 times the normal concentration of growth factors that include PDGF, VEGF, TGF- β locally also accelerating wound healing. Addition of calcium salts also helps in activation of platelets. Skin graft take involves plasma imbibition, inosculation and neovascularization. A successful take of graft depends on wound bed vascular status, micro environment and adequate hemostasis. APRP being rich in platelets augments healing process by platelet plug formation, conversion of fibrinogen to fibrin which in turn helps in adhesion of graft and provides a stable fixation. The growth factors from APRP promote angiogenesis and collagen deposition augmenting take of graft.

CONCLUSION

Autologous platelet rich plasma is an effective measure in enhancing graft take and is a good choice for wound bed preparation in skin grafting provided the patient has a good functional status and surface area to be treated is small.

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Water Surveillance and Surgical Site Infection: A Prospective Study from a Tertiary Care Oncology Centre

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Abstract

Quality of water supply in health-care organisation (HCO) is often not taken seriously in many low to middle income countries (LMIC). Waterborne pathogens may come in contact with patients in various ways in HCOs which includes showering, bathing, drinking or in contact with improperly cleaned medical devices. Hand colonization is another important source of transmission of health-care associated infections (HAIs), specially the Surgical site infections (SSI). Contaminated water system may contribute to HAIs by various pathogens like coliforms and other opportunistic pathogens like *Pseudomonas aeruginosa*, *Burkholderia cepacia*, *Acinetobacter spp.* etc. Stringent implementation of water surveillance (WS) program along with immediate corrective action against the root cause of contamination should be mandatory in each HCO.

Keywords: healthcare organisation (HCO), healthcare associated infections (HAIs), Surgical site infections (SSI), water surveillance (WS), coliforms, *Pseudomonas spp.*

INTRODUCTION

Health care-associated infections (HAI) are the most frequent adverse event in any health-care organization (HCO). The endemic burden of these infections are significantly higher in low-and

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middle-income countries (LMIC) as compared to high-income countries (HIC). In high-income countries, approximately 30% of patients in intensive care units (ICU) are affected by at least one HAI which is 2-3 fold higher in LMIC.¹ In India, the overall prevalence of HAIs was found to be 7%, and surgical site infections (SSI) were the most common (33%).² Another study reveals 1.75 HAI cases per 1000 patient-days; SSIs being predominated HAI, 23.94%.³

Various studies suggest that environmental contamination is associated with HAI by multi-drug-resistant organisms (MDROs) [vancomycin-resistant enterococci (VRE), methicillin-resistant *Staphylococcus aureus* (MRSA), *Acinetobacter baumannii*, *Clostridium difficile*, and *Pseudomonas aeruginosa*], viruses, mycobacteria, and fungi.^{4,5,6}

Contaminated water have been linked to numerous HAI outbreaks which is associated with improper

hand washing and inadequate cleaning of medical devices for reprocessing.⁷ Study on availability of proper environmental conditions and standard precaution items over 78 LMIC including 129,557 HCO showed 50% lacked piped water.⁸ Water supply in HCO are frequently overlooked although it is an essential manageable source to control HAI.⁷

Our study is to evaluate correlation between water condition of the operation theatre (OT) with SSIs that were traced over 2 years in a tertiary care hospital in Delhi along with the actions taken activities.

MATERIAL AND METHODS

This study was carried out in a 200 bedded tertiary care oncology centre over a period of 2 years, from December 2019 to December 2021. Water and SSI surveillance record per month were evaluated.

Water Surveillance (WS) Protocol of Hospital

HCO has formulated a protocol for WS. As per National Accreditation Board for Hospitals and Health Care Providers (NABH) 4th Edition, periodic surveillance activities were appropriately performed for Operation Theatre, which belongs to a high-risk areas (HRA).⁹ One of the surveillance

activities include monthly water culture and quarterly water quality testing.

WATER SAMPLE COLLECTION

Water samples were collected in heat sterilized bottles of 100 ml capacity. Water is allowed to run for 2-3 minutes prior to collection into the bottle.¹⁰ Frequency of water collection for surveillance from OT is once in a month.

Microbiological evaluation: Multiple Tube Test (MTT)¹⁰

MTT was done using MacConkey broth Purple (MCBP) (double strength) with Bromocresol purple (BCP) (HiMediaLabs).¹¹

50 ml and 10 ml volumes of MCBP at double strength and 5 ml at single strength are placed into tubes and bottles containing inverted air free Durham tubes. Then aseptically add 50 ml volume and 10 ml volume of water into bottle/tubes containing corresponding 50 ml and 10 ml volumes of double strength MCBP and five 1 ml volumes into tubes containing 5 ml of single strength medium. [Fig 1] *Escherichia coli* American Type Culture Collection (ATCC) 25922 was used as positive control while *Staphylococcus aureus* ATCC 25923 was used as negative control.



Fig. 1: Multiple Tube Test using MacConkey broth Purple (MCBP) (double strength) with Bromocresol purple (BCP) with positive and negative controls

Bottles and tubes are incubated aerobically at 37 for 24 to 48 hours.

Lactose fermenting organisms (Coliforms) turns the media from blue to yellow. Non-lactose fermenting organisms (example, *Pseudomonas spp.*) do not change the colour but there will be turbidity and pellicle formation in the liquid medium. Positive tubes were further sub-cultured on Blood agar and MacConkey agar for identification, morphology, biochemical testing and antibiogram.

Most probable number (MPN)/ 100 ml of bacteria was derived from McCrady's table used which is based on the numbers of positive and negative reactions in replicate tests of different volumes of the sample examined by the multiple tube method.^{10,12}

Water Quality were Categorized into 4 Groups:¹²

Excellent: 0 MPN/100 ml of water

Satisfactory: 1-3 MPN/100 ml of water

Suspicious: 4-9 MPN/100 ml of water

Unsatisfactory: ≥ 10 MPN/100 ml of water

Surveillance of SSI: ^{13,14}

SSI Data collection form is prepared keeping WHO peri-operative and post-operative data collection forms.¹³ (Fig. 2)

 Dharamshila Narayana Superspecialty Hospital <small>1-800-100-10000 www.dnh.in 011-41666666</small>		SURGICAL SITE INFECTION SURVEILLANCE FORM																					
PATIENT'S NAME: _____ AGE: _____ SEX/M/F: _____ MRN: _____ DOA: _____ WARD: _____ ROOM NO.: _____ CONSULTANT: _____ DATE OF SURGERY: _____ DATE OF DISCHARGE: _____		FINAL DIAGNOSIS: <div style="border: 1px solid black; height: 40px; width: 100%;"></div>																					
SURGICAL PROCEDURE: <div style="border: 1px solid black; height: 40px; width: 100%;"></div>		NUMBER OF PRE-OPERATIVE DAYS IN THE HOSPITAL: _____ FEVER STATUS: <input type="checkbox"/> DAY(S): _____																					
PRE-OPERATIVE DAY: _____ OPERATIVE DAY: _____ 1ST POST-OP DAY: _____ AFTER & CONTINUE: <input type="checkbox"/>		VERBAL COMMUNICATOR: 1. WOUND CLASS: <input type="checkbox"/> 1. CLEAN <input type="checkbox"/> 2. CLEAN-CONTAMINATED <input type="checkbox"/> 3. CONTAMINATED <input type="checkbox"/> 4. DIRTY 5. UNKNOWN 2. ANY KNOWN SOURCES OF CONTAMINATION DURING SURGERY. (PLease tick) EXTERNAL <input type="checkbox"/> <input checked="" type="checkbox"/> BOWEL <input type="checkbox"/> <input checked="" type="checkbox"/> CHEST/BRONCHUS <input type="checkbox"/> <input checked="" type="checkbox"/> URINARY TRACT <input type="checkbox"/> <input checked="" type="checkbox"/> BILARY TRACT <input type="checkbox"/> <input checked="" type="checkbox"/> 3. ANY OTHER CAUSE OF FEVER: VENOUS LUNG'S, URINARY-TRACT, ANYOTHER. 4. DM: <input type="checkbox"/> <input checked="" type="checkbox"/>																					
5. IN YOUR EXAMINATION DOES THE SURGICAL SITE HAVE: ERYTHEMA <input type="checkbox"/> <input checked="" type="checkbox"/> INDURATION <input type="checkbox"/> <input checked="" type="checkbox"/> DISCHARGE <input type="checkbox"/> <input checked="" type="checkbox"/>		IF DISCHARGE IS PRESENT, REBUILT OF INFECTION CULTURE <input type="checkbox"/> <input checked="" type="checkbox"/> GRAM STAIN <input type="checkbox"/> <input checked="" type="checkbox"/>																					
6. PRE-OPERATIVE SHOWER: <input type="checkbox"/> <input checked="" type="checkbox"/> 7. HAIR REMOVAL BY CLIPPER: <input type="checkbox"/> <input checked="" type="checkbox"/> 8. INDUCTION TIME: _____ 9. INCISION TIME: _____ 10. PROPHYLACTIC ANTIBIOTIC TIME (PD): _____ 11. PROPHYLACTIC ANTIBIOTIC: _____ 12. DURATION OF SURGERY: _____ 13. ANY RELOOSE: <input type="checkbox"/> <input checked="" type="checkbox"/>																							
14. CARE COMPANION PROGRAMME (CCP): <input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> <input checked="" type="checkbox"/> 15. POST-OP DRESSING IN-HOUSE: <input type="checkbox"/> <input checked="" type="checkbox"/>																							
16. SHOWER AFTER SURGERY: <input type="checkbox"/>																							
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SIGNATURE OF ICO/CONSULTANT SURGEON ON SURGICAL WOUND CLASSIFICATIONS HOSPITAL SIGNATURE 1. CLEAN: <ul style="list-style-type: none"> • Intact surgical incision • Clean, no GI tract entered • Clean-contaminated • Sterile, no foreign materials, neck dissection, thyroid, muscle, heart, skeleton 2. CLEAN-CONTAMINATED: <ul style="list-style-type: none"> • Clean, GI tract entered, controlled • CONTAMINATED: • Enteric, GI, skin, multiple, loose tissue, parenteral surgery, bronch, colon surgery 3. CONTAMINATED: <ul style="list-style-type: none"> • Major tissue or muscle wounds • Major tissue or muscle technique • Gross spillage from GI tract • Enteric, GI, skin, multiple, loose tissue, parenteral surgery, bronch, colon surgery • Exogenous infected body site syndrome (e.g., dialysis, medical device, medical surgery, penetrating wound) 4. DIRTY: <ul style="list-style-type: none"> • Contaminated wounds • Gastrointestinal perforation • Organisms present before surgery (e.g., <i>Escherichia coli</i> & <i>Staphylococcus aureus</i>, peritonitis, wound dehiscence) 																							

Fig. 2: Surveillance form for Surgical site infection

Incidence of SSI=Number of SSI cases detected during the surveillance period x 100/Number of total surgical patients during the surveillance period

Follow up period for SSI was considered as 30 days (eg. Neck surgery, gastric surgery etc) and 90 days (eg. Cardiac surgery, Breast surgery etc.) as per CDC guideline. The wounds were categorized into four types: Clean (C), Clean-Contaminated (CC), Contaminated (CO), and Dirty/Infected (D).¹⁴

As per World Health Organization (WHO), the pooled SSI incidence was 5.6 per 100 surgical procedures (95% CI: 2.9–10.5).¹⁵ In our HCO, average rate of SSI is 1.95 (95% Confidence Interval 1.74 to 2.63).

RESULTS:

Overall HAI in our HCO over 2 years (December 2019 to November 2021) were found to be 82; out of which SSI predominated with 57% (47/82) (Fig. 3)

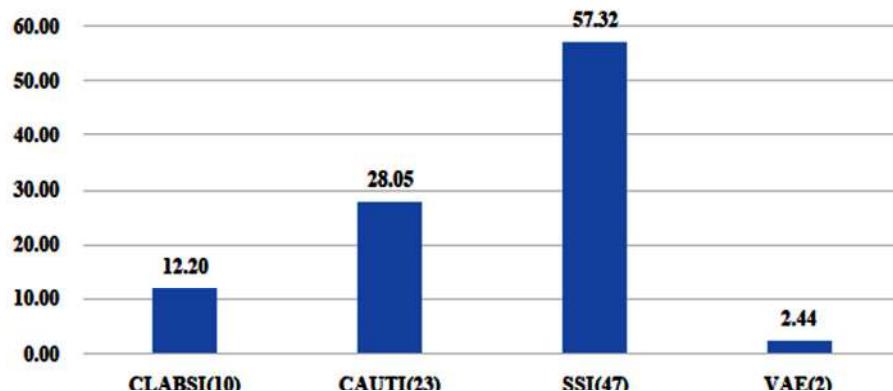


Fig. 3: Distribution of Hospital Acquired Infections (%) over 2 years

Note: CLABSI:Central line associated blood stream infection; CAUTI: Catheter associated urinary tract infection; SSI: Surgical site infection; VAE: Ventilator associated pneumonia

Out of all SSIs, 78.72% (37/47) were clean contaminated wound, 19.15% (9/47) were contaminated wound. (Fig 4a) Majority of the SSIs were related to oncology surgeries, 93.62%(44/47). [Fig 4b]

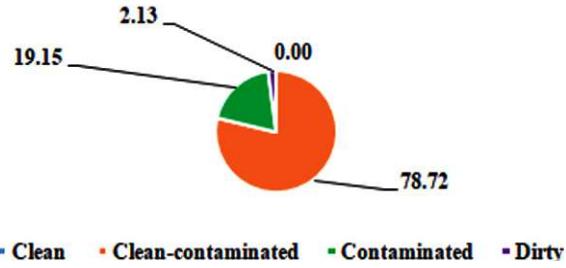


Fig. 4a: Percentage of distribution of Surgical Site Infections (n=47) as per classification

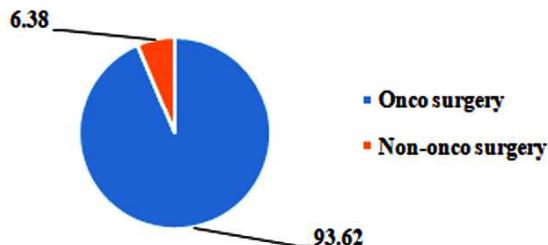


Fig. 4b: Percentage of distribution of Surgical Site Infections (n=47) as per Oncology or Non-oncology.

WS in OT showed excellent water quality with 0 MPN/100 ml in 62.50%, suspicious water quality with 4-10 MPN/100 ml in 4.17% and unsatisfactory water quality with ≥ 10 MPN/100 ml in 33.33% over 2 years. (Fig. 5)

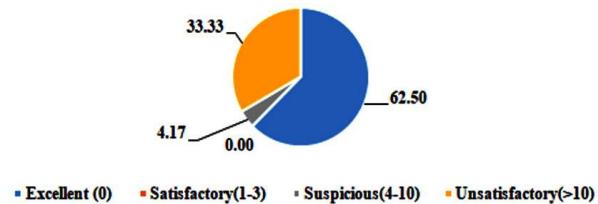


Fig. 5: Rate of distribution of water quality in Surveillance Water Culture as per MPN/100 ml



Fig. 6: Pall Aquasafe Water filters installation

Surveillance of SSI over 2 years (December 2019 to November 2021) showed highest rate of SSI in October 2020 (4.95%) followed by August 2020 (3.87%). (Table 1)

Recursive partitioning analysis was carried out using rpart package¹⁶ in R¹⁷ to generate optimal cut off for SSI which was found to be 1.95 over these 2 years (Dec-19 to Nov-21) which was considered as benchmark for SSI in our institute.

Considering 1.95 as benchmark for SSI, relation of growth in WS and number of SSI were established shown in Fig 5. There is statistically significant correlation between SSI rate >1.95 with water contamination with p value 0.0147. (Table 1)

Table 1: Distribution of Surgical Site Infections Over 2 years Along with Result of Water Surveillance (WS) in OT

Months	No. of SSI	Total No. of surgeries	SSI (%)	OT Water Surveillance culture (Growth=1; No growth=0)
Dec 19	3	165	1.82	0
Jan 20	2	154	1.30	0
Feb 20	1	99	1.01	0
March 20	1	51	1.96	0
April 20	0	146	0.00	0
May 20	1	105	0.95	0
June 20	2	137	1.46	1
July 20	2	132	1.52	0
August 20	6	155	3.87	1
Sept 20	2	144	1.39	1
Oct 20	5	101	4.95	1
Nov 20	2	143	1.40	0
Dec 20	1	158	0.63	0
Jan 21	0	169	0.00	0
Feb 21	0	166	0.00	0
March 21	0	110	0.00	1
April 21	2	101	1.98	1
May 21	1	157	0.64	0
June 21	4	199	2.01	1
July 21	5	203	2.46	1
August 21	1	210	0.48	0
Sept 21	1	231	0.43	1
Oct 21	4	207	1.93	0
Nov 21	1	207	0.48	0

Water culture growth and growth in SSI are correlated by phenotypic method and antibiogram. *Pseudomonas spp.* was the predominating isolate in

water culture with considerable MPN/100 ml. Type of growth in WS and similar isolates retrieved from SSI were compared. Species with similar phenotype

and antibiogram were considered having common source of infection. (Tab 2)

Table 2: Correlation of Growth in water culture and Growth in SSI (Phenotyping and Antibiogram)

Months	Organism in water culture (>180 MPN/100 ml)	No. of SSI	Same organism in SSI (No.)
June 20	<i>Pseudomonas</i> spp.	2	<i>Pseudomonas</i> spp. (1)
August 20	<i>Pseudomonas</i> spp.	6	<i>Pseudomonas</i> spp. (1)
Sept 20	<i>Pseudomonas</i> spp.	2	nil
Oct 20	<i>E.coli</i> (faecal coliform)	5	<i>E.coli</i> (1)
March 21	<i>Pseudomonas</i> spp.	0	nil
April 21	<i>Pseudomonas</i> spp.	2	nil
June 21	<i>Pseudomonas</i> spp. and <i>Klebsiella pneumoniae</i>	4	<i>Pseudomonas</i> spp. (1) <i>Klebsiella pneumoniae</i> (2)
July 21	<i>Pseudomonas</i> spp.	5	<i>Pseudomonas</i> spp. (1)
Sept 21	<i>Pseudomonas</i> spp.	1	nil

ACTIONS TAKEN

- **Inter-laboratory comparison:** Inter-laboratory comparison of water testing is done from National Accreditation Board for Testing and Calibration Laboratories (NABL) accredited laboratories which not only includes water culture but also checks various other parameters of water [pH, Total Dissolved Solids (TDS), General parameters like aluminium, ammonia, boron, calcium etc; Toxic parameters like cyanide, lead etc. and conductivity]. No discrepancy was observed in culture results.
- **Local chlorination of the tanks:** Chlorination is done on daily basis in the supply water of the hospital. Daily monitoring is done for TDS, Ph, Free chlorine (Ionexchange, Indion). Residual chlorine level is targetted to keep in between 0.2-0.05 mg/L.¹⁸
- **Cleaning of the tanks:** For mechanized dewatering, a portable submersible was used which used to empty the tank with the flow rate 30,000 Ltrs/hour. After dewatering, dirty water and sludge are removed with the help of a sludge pump. This is followed by cleaning of the walls and ceiling of the tanks with high-pressure jet, which dislodges the layer of dirt and algae. After cleaning of walls & roof, the floor of the tank is thoroughly cleaned by the high pressure water jet removing all the remaining dirt and algae. After removal of the sludge, dirt is vacuumed out using an Industrial Vacuum Cleaner. The sludge is then taken out of the tank and disposed off to a safe place. The same process is adapted for the overhead terrace tanks as well. In the final stage of the tank cleaning, UV Radiation is applied inside the tanks.
- Frequency of cleaning of the tanks are defined for particular area of the hospital. However, whenever there is growth in WS, frequency of cleaning is increased and actions taken promptly.
- **Pall-Aquasafe water filter installation:** As a temporary solution, Pall Aquasafe Water filters (AQ14F1R) were installed. The sterilizing membrane is validated at 0.2 µm and protects against waterborne particulates and pathogens as *Legionella* spp., *Pseudomonas* spp. and fungi. These filters are compatible with upto 1.0 mg/L of ClO₂ (1 ppm) at 60 or with 100 ppm free chlorine at ambient temperature.¹⁹
- **Pipeline change:** Finally pipelines were changed in OT when repeated cultures were showing growth despite all measures.

DISCUSSION

SSI accounts for 20% of all HAIs with 9-11 fold increased risk for mortality despite having advanced operating room ventilation, sterilization methods, barriers, surgical technique, and availability of antimicrobial prophylaxis.²⁰ Our study reveals 57% SSI out of all HAIs which is higher as compared to other literature in India (33%)², 20%²⁰; this may be because SSI rates are not routinely documented in large number of HCOs all over the LMIC; official data on SSI is just the tip of the iceberg. Secondly out of all SSIs, 93.62% belonged to surgeries on immunocompromised cancer patients who are more prone for such

infections. Moreover, surgeries on such patients were mostly categorized into clean-contaminated and contaminated surgeries keeping the risk of SSI on a higher side.(Fig 3a, 3b) The development of SSI is multi-factorial; it may be related to patient's risk factors such as age, sex, co-morbidity, smoking habit, obesity, nutrition level, immunosuppression, malignancies, transfusion etc. Environmental risk factors include level of microbial contamination, temperature, humidity, air renewal etc.^{21,22}

There are evidences since long that microbes can survive in hospital water reservoir; various studies have confirmed hospital water as a source of nosocomial infection.^{7,23,24} Literature suggests relation of damp environmental reservoirs (sink drains, traps and the horizontal drainage system) with Multi-drug resistance (MDR) Gram-negative bacilli, including MDR coliforms.^{21,25} *K. pneumoniae* demonstrating prolonged survival within plumbing components are also more likely to be extended spectrum β -lactamases (ESBL) producers.²⁶ There are many articles in favour of MDR *K. pneumoniae* persistently harboring in sink and related pipes leading to outbreaks in ICUs.^{21,27,28,29} In LMICs, inadequate chlorination is associated with high levels of water contamination which are linked to outbreaks by Enterobacteriaceae, including *Klebsiella spp.* and *Enterobacter spp.*⁷ In our study, we found MDR-*Klebsiella pneumoniae* once in WS; 2 cases of SSI in that month was also shown growth of MDR-*Klebsiella pneumoniae*. *E. coli* is a type of fecal coliform bacteria commonly found in the intestines of animals and humans and the presence of it in water is a strong indication of recent sewage or animal waste contamination.³⁰ In our HCO, faecal *E. coli* was isolated once which was immediately taken care of. However, out of 5 cases of SSI, one had *E. coli* infection with same susceptibility pattern with that of isolate in water. No *Enterobacter spp.* was isolated in WS.

Coliforms are known as the best index for monitoring water microbial quality; however, growth of coliforms can be inhibited by heterotrophic bacteria. Hence, as an alternative index in water microbial quality control, *Pseudomonas spp.* can be taken as one indicator.^{31,23} Water of HCO can act as a reservoir for opportunistic pathogens like *Pseudomonas spp.*, *Stenotrophomonas spp.*, *Burkholderia cepacia*, *Acinetobacter spp.*, fungi, etc. which pose a risk of colonization and infection for immunocompromised patients.^{21,32} These organisms are known for biofilm formation on sinks, sink traps, pipes, water lines and hospital drains. These organisms residing in biofilms are resistant

to the effect of chlorine and other disinfectants and antibiotics leading to Antimicrobial resistance (AMR).^{20,32} Physical disruption of the biofilm lining the internal surfaces of affected water systems is the reliable method of cleaning.^{21,34,35} Water related *Pseudomonas spp.* was the notorious microorganism in our study which was found in many SSIs simultaneously.

Modes of transmission includes direct contact, ingestion of water, indirect contact, inhalation of aerosols dispersed from water sources, and aspiration of contaminated water.^{23,36} High levels of bacteria in hospital water, sinks, faucets, or shower heads has been associated with hand colonization.¹⁸ Although there is no standardized limit of bacteriological clean water for surgical hand scrub, surgical hand antisepsis requires medicated soap with clean water to rinse the hands.³² Thereafter, micro-perforations in the surgical gloves can lead to transmission of colonized microorganisms from the hands of the surgeon to the patient that happens at an average of 18% (5-82%) at the end of the surgery.^{32,37} Double gloving decreases the risk of puncture during surgery, but punctures are still observed in 4% of cases after the procedure.^{32,38} Therefore, continuous monitoring and surveillance of water quality is a real need in any HCO, with special reference to high risk areas like OT. Primarily being a cancer care organization along with transplants, we can't ignore any microorganism in WS culture from OT; for which various strategic multi-level actions were taken as soon as possible to reduce the rate of SSI.

CONCLUSION

Contaminated water system may contribute to HAIs by various pathogens like coliforms and other opportunistic pathogens. Stringent implementation of WS program along with immediate corrective action against the root cause of contamination should be mandatory in each HCO. *Pseudomonas spp.*, which causes opportunistic infections in immunocompromised patients was found to be the predominating resistant bug in our HCO. This bug was resistant to all cleaning efforts; finally pipelines had to be changed to get rid of this resistant organism along with stringent protocols of chlorination and tank cleaning. Re-contamination is not uncommon despite every effort; therefore routine policies are made for periodic surveillance of water with defined frequency in each high risk area of the HCO.

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A Study of Malaria in Lakshadweep Islands

Arvind Nath

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Abstract

Background: To the best knowledge of the author, there is no available literature on the status of Malaria in Lakshadweep Islands. Hence the preparation of this document.

Objectives: To find out the parameters of Malaria in Lakshadweep Islands till as recently as possible.

Methods: By studying the documents prepared by the National Centre for Vector Borne Diseases (NCVBD) and the website of Department of Medical & Health Services, Lakshadweep Islands.

Results: It is seen that the Annual Parasite Incidence (API) of Malaria in Lakshadweep Islands was at the low value of 0.08 in 2018 and there were no indigenous cases of Malaria in the islands. All the cases of Malaria seen there were imported cases.

Conclusions: Lakshadweep Islands may be identified as having achieved Malaria-elimination goals.

Keywords: Malaria, Lakshadweep Islands, API, NCVBD, WHO

INTRODUCTION

Lakshadweep Islands lie towards the south-western part of the Indian mainland. They are bounded by the Arabian Sea in the west and north, the Indian Ocean in the south and the Lakshadweep Sea in the east.

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Lakshadweep Islands lie towards the south-western part of the Indian mainland. They are bounded by the Arabian Sea in the west and north, the Indian Ocean in the south and the Lakshadweep Sea in the east.

Anti-Malaria activities are carried out throughout the Lakshadweep Islands. While six percent of the total population is covered for active surveillance, four percent is monitored for passive surveillance each year. Blood samples are collected from all the fever cases and from contacts and during surveys. The Malaria positive cases are diagnosed and treated. Focal spraying with insecticides and observance of Anti-Malaria month during June to create awareness among the people are other measures taken.²

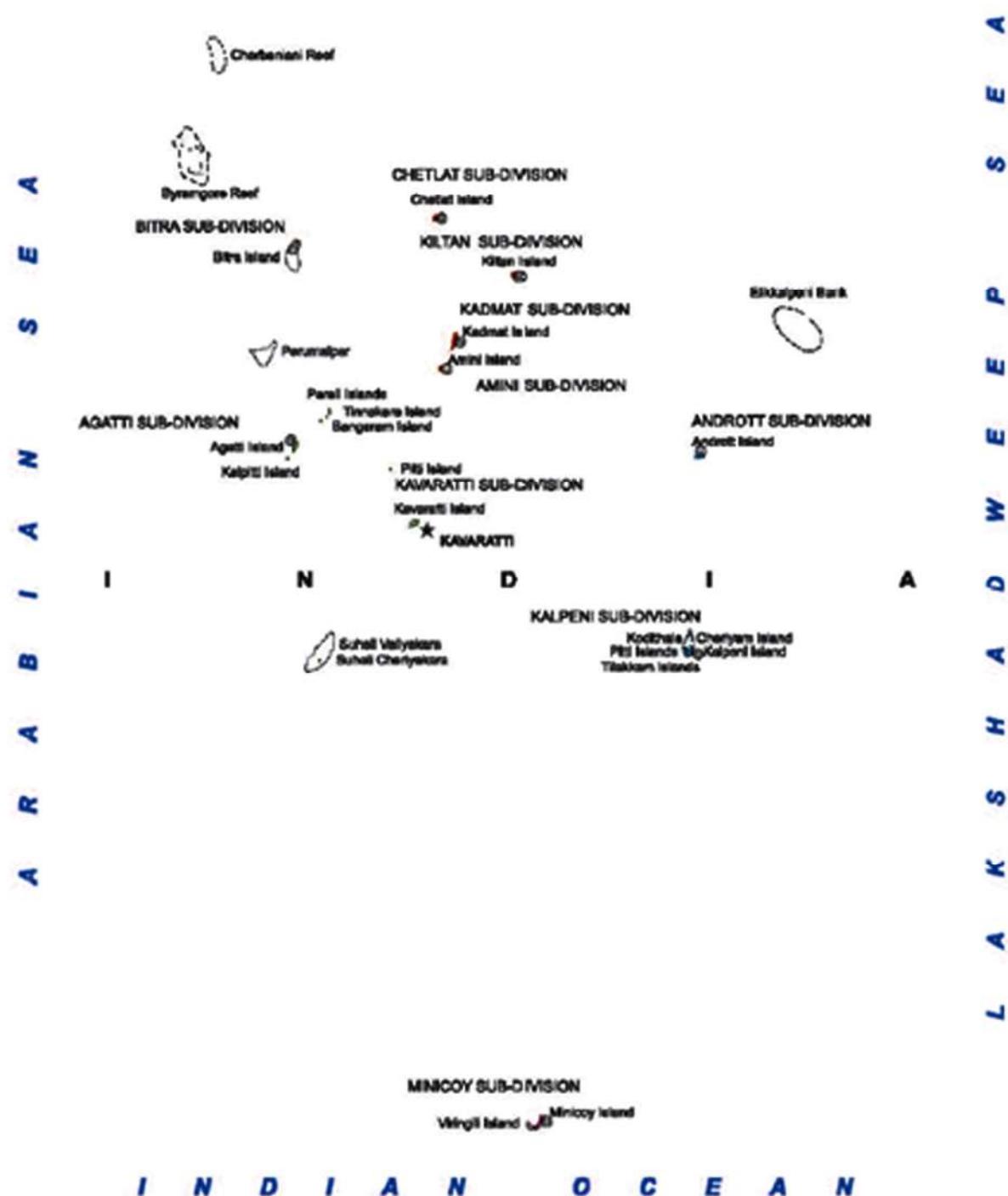


Fig. 1: Map of Lakshadweep Islands [Source: (1)]

MATERIAL & METHODS

The study design included analysis of the annual reports of the Malaria Division of the National

Centre for Vector Borne Diseases Control (NCVBDC) for 2017 and 2018 and a study of the website of Department of Medical & Health Services, Lakshadweep Islands.

RESULTS

According to the most recent data available on the NCVBDC website (data for 2018), the API for Lakshadweep Islands was 0.08.² It's comparison with the API from 2017 can be seen from the following table:

Thus, it is seen that the API in 2018 was four times as that seen in 2017.

Table 1: API of Lakshadweep Islands, 2017 and 2018

Union Territory	Year	
	2017	2018
Lakshadweep Islands	0.02	0.08

Sources: (2) and (3)

A study of the website of Department of Medical & Health Services, Lakshadweep Islands revealed that Malaria is totally controlled except for a few imported cases.⁴

DISCUSSION

Indigenous Malaria does not exist in Lakshadweep Islands. The Malaria which occurs there is due to imported cases. However, as seen from the data, the rate of importation seems to be rising.

In 2016, the Government of India adopted a framework for Malaria Elimination in India covering the period 2016–2030.⁵ This was based on WHO's Global Technical Strategy for Malaria covering the period 2016 – 2030 which was adopted in 2015 and updated in 2021.⁶

The aim is to reach zero Malaria cases by 2027 and then wait for three years before WHO can grant Malaria-free status certification. It is already nearly the middle of 2022 and India is about to reach the halfway mark of the period from 2016 to 2027.

CONCLUSION

Although Lakshadweep Islands did not reach zero Malaria cases in 2018, whatever disease is occurring is due to imported cases. Therefore, it is a good candidate for being the first administrative jurisdiction in the country to be able to achieve Malaria elimination goals.

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